Forms of cytochrome oxidase

OKUNUKI and his colleagues^{1,2} have recently reported that the spectrum of a cytochrome oxidase preparation shows some differences according to whether it is oxidized by $K_3Fe(CN)_6$, or by O_2 , and have suggested that the spectrum obtained with oxygen is due to formation of an O_2 complex of the same type as oxyhaemoglobin. As already communicated to the Fourth International Congress of Biochemistry in September, 1958 we have also observed a difference between the two spectra but our studies make it doubtful that the compound formed with O_2 is a complex with molecular O_2 .

The SMITH AND STOTZ³ "type II" preparation of cytochrome oxidase was used. It contained cytochromes a and a_3 , and was free from spectroscopically identifiable amounts of cytochromes b, c and c_1 , and of mitochrome⁴.

Fig. 1 shows the absorption spectra between 576 and 641 m μ under four different conditions. Curve 1 is the spectrum of the preparation in the presence of air. Curve 2 was obtained 18 h after evacuation of a solution. Under these conditions, cytochromes a and a_3 are slowly and simultaneously reduced, the degree of reduction reaching 95% of that obtained with Na₂S₂O₄ (curve 3). When the evacuated solution was opened to air and O₂ bubbled through, the spectrum shown in curve 1 was slowly restored.

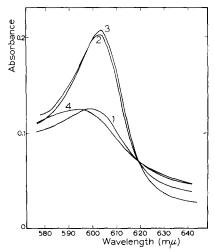


Fig. 1. Spectra of cytochrome oxidase. Curve 1 was obtained in air, curve 2 after evacuation of a solution, curve 3 with $\mathrm{Na_2S_2O_4}$ in $\mathrm{N_2}$, and curve 4 by addition of $\mathrm{K_3Fe(CN)_6}$ 2.4·10⁻⁴ M or 2.4·10⁻³ M) to the evacuated solution.

Curve 4 shows the spectrum obtained by addition of $K_3Fe(CN)_6$. The same spectrum was obtained whether the $K_3Fe(CN)_6$ was added in the presence of air or anaerobically to a preparation reduced spontaneously by evacuation. Dialysis of the $K_3Fe(CN)_6$ -treated preparation did not change the spectrum. The cytochrome oxidase activity (measured manometrically with cytochrome c and ascorbic acid) was not affected by treatment with $K_3Fe(CN)_6$, followed by dialysis.

Calculations showed that the spectrum of curve I could not be obtained by addition of curves 3 and 4 in various proportions. Thus, it cannot be due to the

Abbreviations: $a \cdots$, $a_3 \cdots$, ferricytochrome a and a_3 , $a \cdots$, $a_3 \cdots$, ferrocytochrome a and $a_3 \cdots$

presence of an inactive cytochrome a or a₃ which is oxidizable by K₃Fe(CN)₆ but not by O2, as suggested by Chance1 to explain Okunuki's results. A compound not represented in curves 3 and 4 must be at least partly responsible for the spectrum

The positions of the peaks of the absorption bands (in air, 420 and 598 mu; with $Na_2S_2O_4$, 444 and 605 m μ ; with $K_3Fe(CN)_8$, 424 and 591-3 m μ) differ in some respects from those given by Okunuki^{1, 2}, but they agree in showing that the spectrum of cytochrome $(a + a_3)$, oxidized with $K_3Fe(CN)_6$, is not the same as that obtained with O_2 . It seems probable that ferricyanide gives the combined spectrum of $a \cdots$ and $a_3 \cdots$. Since O_2 reacts with $a_3 \cdots$, curve 1 probably represents the combined spectrum of $a \cdots$ and a second oxidized form of cytochrome a_3 . Our experiments make it unlikely that the latter is a complex of molecular O_2 and a_3 ., since both the formation of the spectrum of $(a \cdot \cdot + a_3 \cdot \cdot)$ on evacuation, and its disappearance on re-oxygenation, were very slow reactions. It is possible, however, that the second oxidized form contains an oxygen atom bound to the ferrous iron atom, as in ferryl compounds5.

Although it is not yet known whether both forms of oxidized cytochrome a_3 are concerned in the enzymic reaction, it is significant that both are equally active. Since it is likely that a_3 ·· is concerned in the enzymic reaction, both forms of oxidized cytochrome a_3 must be reducible in the presence of ascorbic acid and cytochrome c.

In summary, it is suggested that curves 3, 4 and 1 given in Fig. 1 represent the spectra of $a \cdot \cdot + a_3 \cdot \cdot \cdot$, $a \cdot \cdot \cdot + a_3 \cdot \cdot \cdot$ and $a \cdot \cdot \cdot$ and $a_3 \cdot \cdot \cdot$ O (or another form of oxidized cytochrome a_3), respectively. The interconversions of the three forms of cytochrome a_3 are possibly described by the following scheme

$$\mathbf{a_3} \cdots \underbrace{ \begin{array}{c} \mathbf{K_3Fe(CN)_6} \\ \\ \mathbf{A_3} \cdots \end{array}}_{\mathbf{A_3}} \mathbf{a_3} \cdots \underbrace{ \begin{array}{c} \mathbf{O_2} \\ \hline \mathbf{AH} \end{array}}_{\mathbf{a_3}} \cdots \mathbf{O}$$

where AH represents reducing compounds present in the enzyme preparation. The presence of ascorbic acid and cytochrome c in the manometric method of measuring the cytochrome oxidase reaction may cause an increase in the rate of some of these reactions. Additional reactions may also be introduced (cf. SLATER⁶). In this connection, the report of Sekuzu et al.2 that, in the presence of cytochrome c, O2 gives the same compound as obtained with ferricyanide (i.e. $a_3 \cdots$ in the above scheme) is of great interest.

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